

## Characterization of the NanoSpark® GROW-NK Soluble Activator, a Feeder Cell, and Magnetic Bead-Free Natural Killer Cell Expansion Technology

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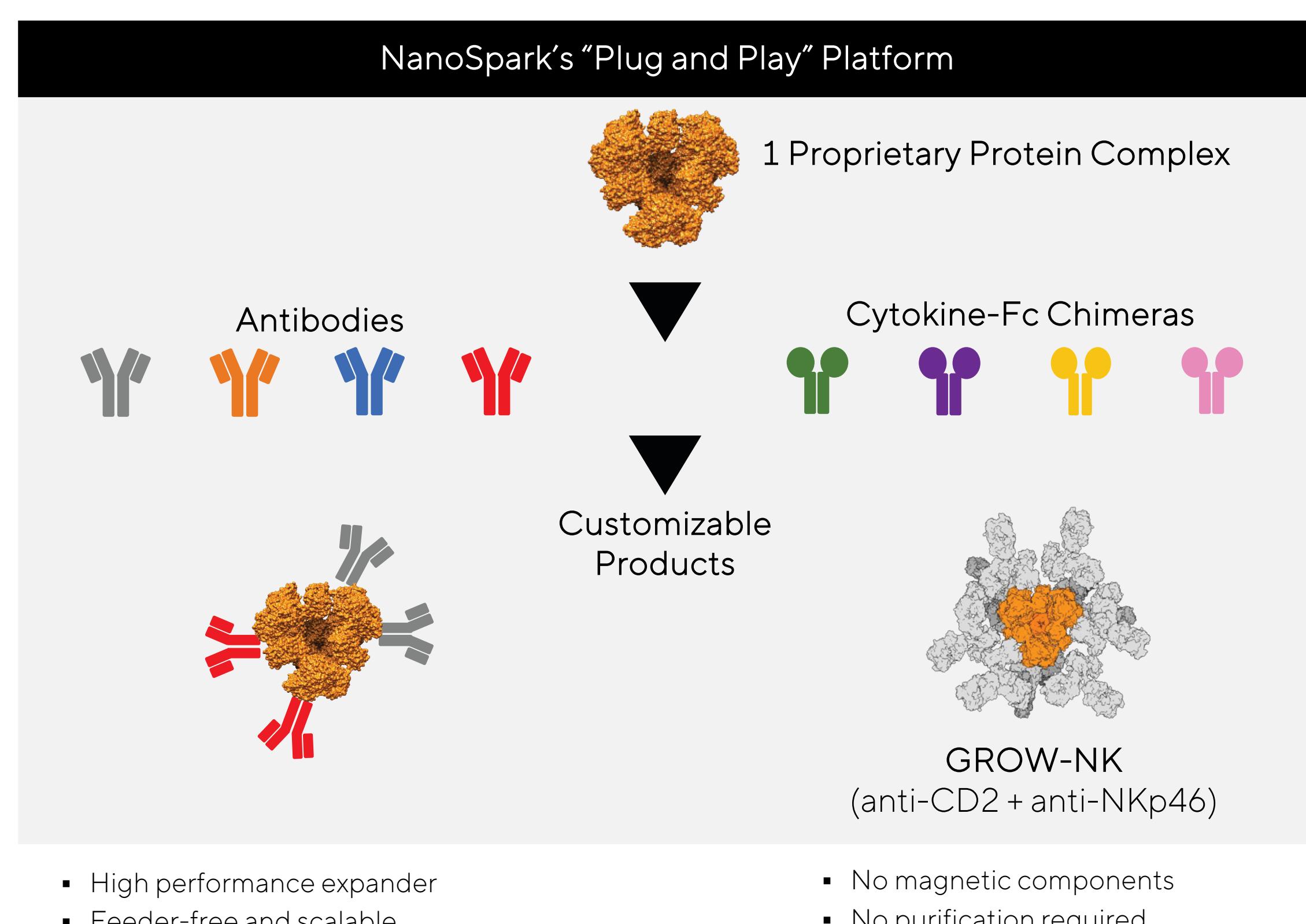
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### Introduction

Natural killer (NK) cells have generated strong interest in the cell therapy field for combating various types of cancers. NK cells offer unique advantages to this emerging industry: they require no HLA matching and are excellent candidates for “off-the-shelf” therapies. NK cells can identify and attack cancer cells even without prior stimulation. However, a major limitation of NK cell therapies is the challenge of producing a sufficient quantity of NK cells to achieve a therapeutic effect. The current industry standard for NK cell expansion involves co-culturing NK cells with engineered feeder cell lines. These feeder cells are costly, labor-intensive to maintain, and require purification—a process that is time-consuming and difficult. Even after purification, feeder cells can still contaminate the final cell product. Nanotein’s novel platform technology, NanoSpark®, helps overcome these obstacles.



### Experimental Approach

- Figure 1 Cryopreserved peripheral blood (PB) NK cells were expanded in CellGenix® GMP SCGM medium with IL-2/IL-15 (35 ng/mL each), HPL, and NanoSpark® GROW-NK (20  $\mu$ L/mL). Cultures were maintained up to 55 days and analyzed for CD3-CD16+CD56+ by flow cytometry<sup>1</sup>.
- Figure 2. Cryopreserved PB NK cells were cultured in CellGenix® GMP SCGM medium with 5% human serum and various cytokine combinations (IL-2/IL-15: 35 ng/mL each or IL-2/IL-15/IL-21: IL-2, IL-15, 35 ng/mL each, and IL-21, 10 ng/mL). NanoSpark® GROW-NK (1-20  $\mu$ L/mL) was added with or without weekly restimulation. NK phenotype (CD3-CD16+CD56+NKp46+) and cytotoxicity (CD3-CD16+CD107a+, day 28-expanded NK effector cells were co-incubated with K562 target cells at effector-to-target (E:T) ratios 1:1, 5:1, 10:1, 4 h) were assessed by flow cytometry<sup>2</sup>.
- Figure 3. Cryopreserved PB NK cells were expanded in CellGenix SCGM GMP medium supplemented with 5% human serum, IL-2 (20 ng/mL), with or without IL-15 (20 ng/mL), and IL-21 (10 ng/mL), and 5  $\mu$ L/mL NanoSpark® GROW-NK (restimulated twice weekly). Co-cultures with feeder cells K562-mbIL21/41BBL served as controls. Cytotoxicity was tested at E:T 5:1 after 24 h.



Figure 1: GROW-NK Enables Rapid and Long-Term Expansion of Peripheral Blood NKs Under Serum-Free, Feeder-Free Conditions

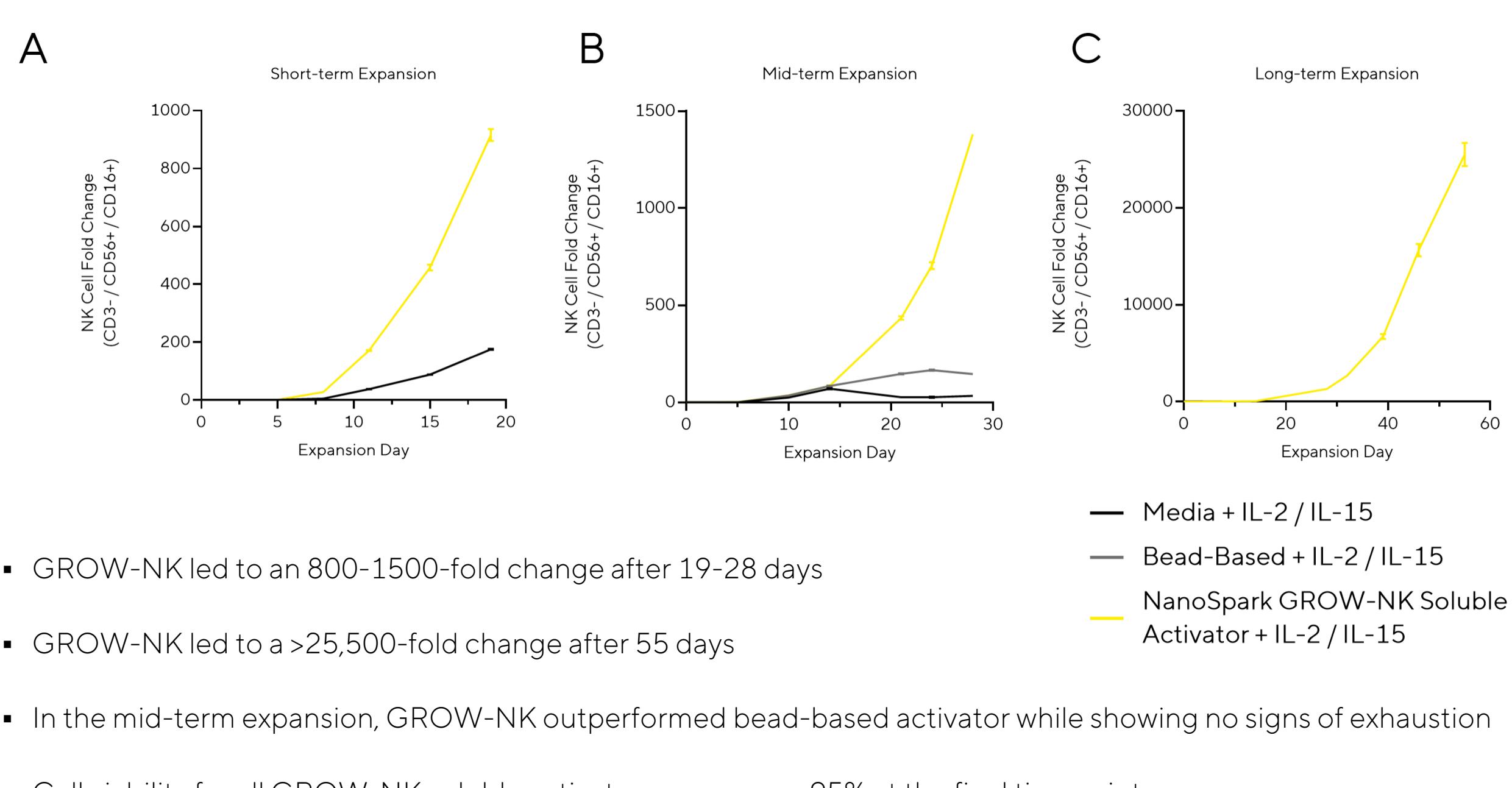
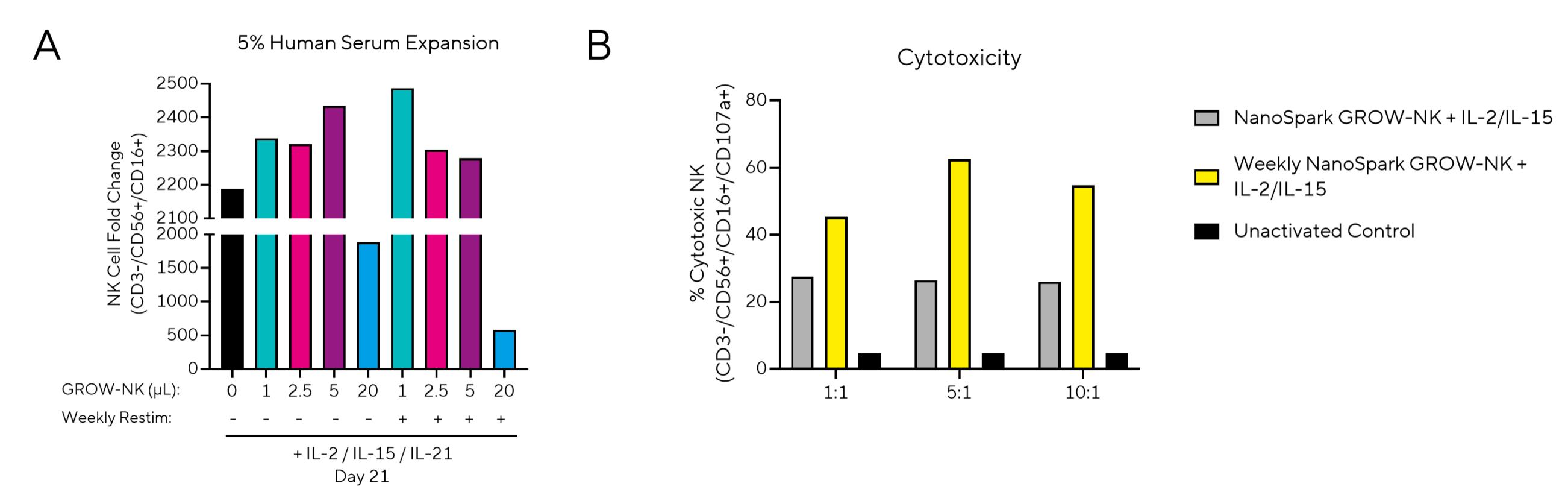
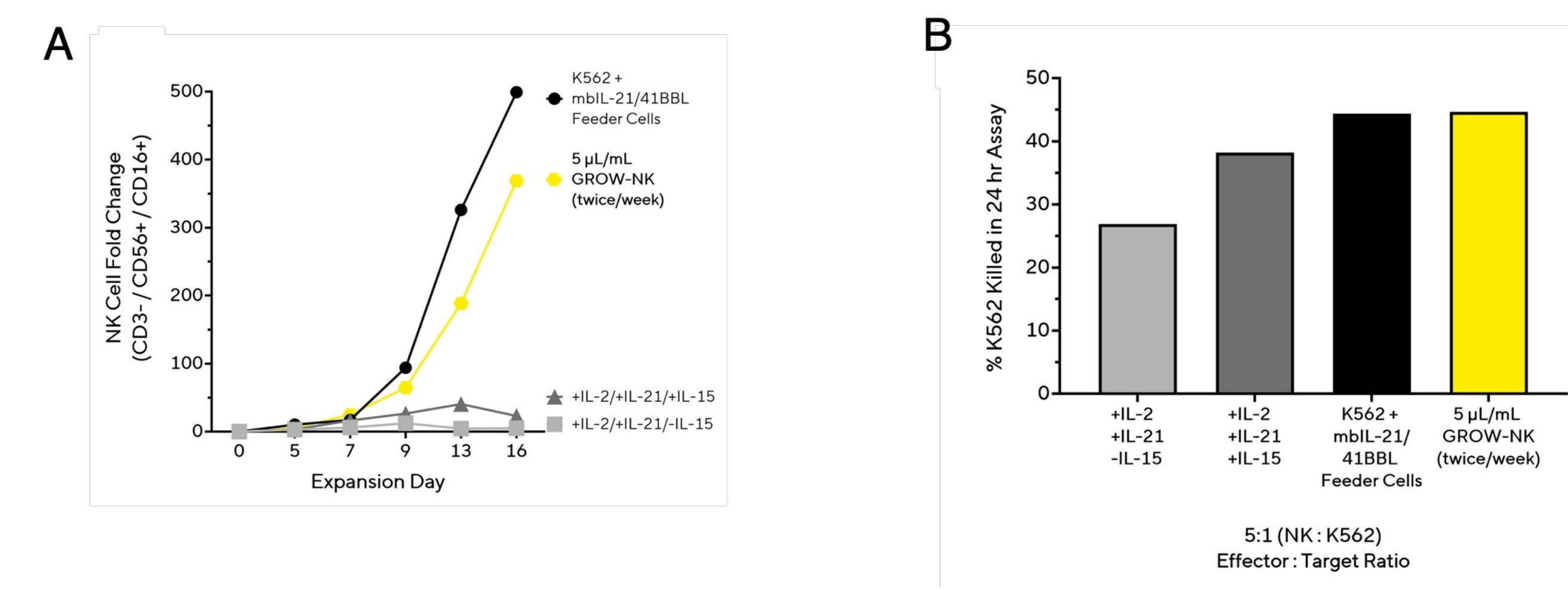


Figure 2: Optimizing GROW-NK Dosing for Maximal NK Expansion and Cytotoxicity Under Feeder-Free Conditions with 5% Human Serum



- A single dose of 5  $\mu$ L/mL GROW-NK resulted in the highest fold expansion (2,433-fold), with the optimal range observed between 1-5  $\mu$ L/mL
- Weekly dosing at 1  $\mu$ L/mL achieved the highest fold expansion overall (2,486-fold), also within the 1-5  $\mu$ L/mL range
- Weekly GROW-NK treatment led to enhanced cytotoxicity compared to a single-dose regimen

Figure 3: Feeder-Free NK Expansion Using GROW-NK Achieves Equivalent Cytotoxicity and Proliferation



- Cells cultured with GROW-NK exhibited comparable fold expansion compared to those co-cultured with engineered feeder cells
- Equivalent killing functionality was observed in GROW-NK groups compared to those co-cultured with engineered feeder cells
- GROW-NK and feeder groups both had >95% viability at the final time point of the expansion

### Conclusion

- Our findings demonstrate a new way to expand NK cells without the need for feeder cells, bead-based activators, or magnetic components
- GROW-NK soluble activator has a comparable expansion and killing profile compared to engineered feeder cells – without any post-expansion purification needed
- This novel, entirely protein-based, modality represents a significant advancement in the activation and expansion of NK cells at scale, paving the way for improved scalable manufacturing for cell therapies against various types of cancers

### References

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2. Aktas, E., Kucuksezer, U. C., Bilgic, S., Erten, G., & Deniz, G. (2009). Relationship between CD107a expression and cytotoxic activity. *Cellular immunology*, 254(2), 149-154.

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