

iQue[®] Antibody Internalization Reagents

- iQue[®] Human Antibody Internalization Reagent
- iQue[®] Mouse Antibody Internalization Reagent

Product Information

Open immediately upon arrival and store reagents at temperatures stated on labels. For research use only.

Product Name	Cat. No.	Format
iQue [®] Human Antibody Internalization Reagent		
	90564	1 × 96-well
	90565	1 × 384-well
iQue [®] Mouse Antibody Internalization Reagent		
	90566	1 × 96-well
	90567	1 × 384-well

Safety data sheet (SDS) information can be found on our website at www.sartorius.com

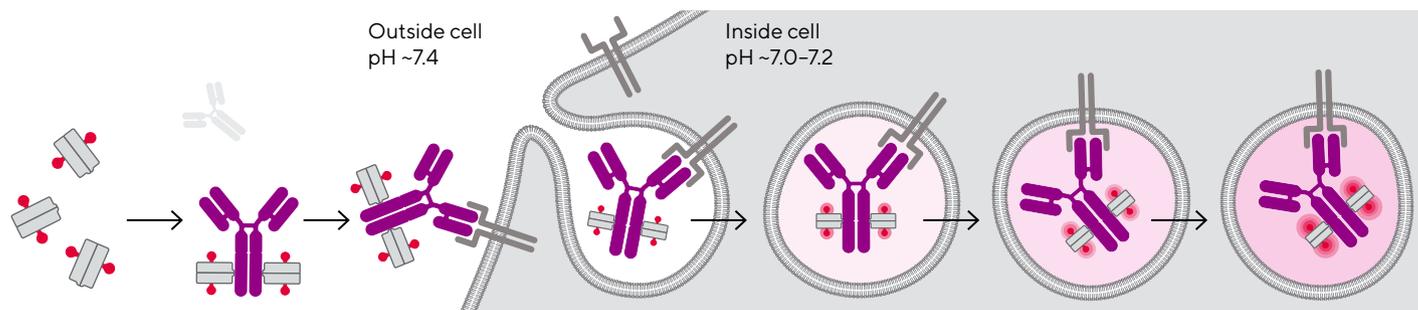
Introduction

Antibody internalization and associated kinetics are critical characteristics that impact antibody therapeutics, pharmacokinetic profiles, and influence the mechanism of action of antibody drugs. For example, antibody-drug conjugates may require rapid internalization to deliver their payload directly into cells to avoid systemic dissemination. However, rapid antibody internalization can be detrimental to immune-mediated killing mechanisms such as antibody-dependent cell cytotoxicity or complement-dependent cytotoxicity.

During biologic drug discovery, antibodies are engineered to increase their biological potencies, to optimize conjugations to non-protein components, and to increase serum half-life. These modifications can have profound effects on antibody activity and specificity, as well as antibody internalization. This necessitates thorough *in vitro*, high throughput screening of antibodies during the early discovery process to quickly identify the most suitable drug candidates for further development.

iQue® Antibody Internalization Reagent is a novel, pH sensitive dye that identifies antibody internalization in a simple plate-based format. The mouse and human types are designed for use with antibodies containing either mouse or human Fc regions. The assay features the flexibility to combine other validated reagents for multiplexed, no-wash protocols with high throughput capabilities. The ability to quickly profile and compare large sets of antibodies and characterize their key attributes, such as antibody internalization, can vastly reduce the time required for candidate generation and expedite the development of potential therapeutic treatments.

Assay Principles

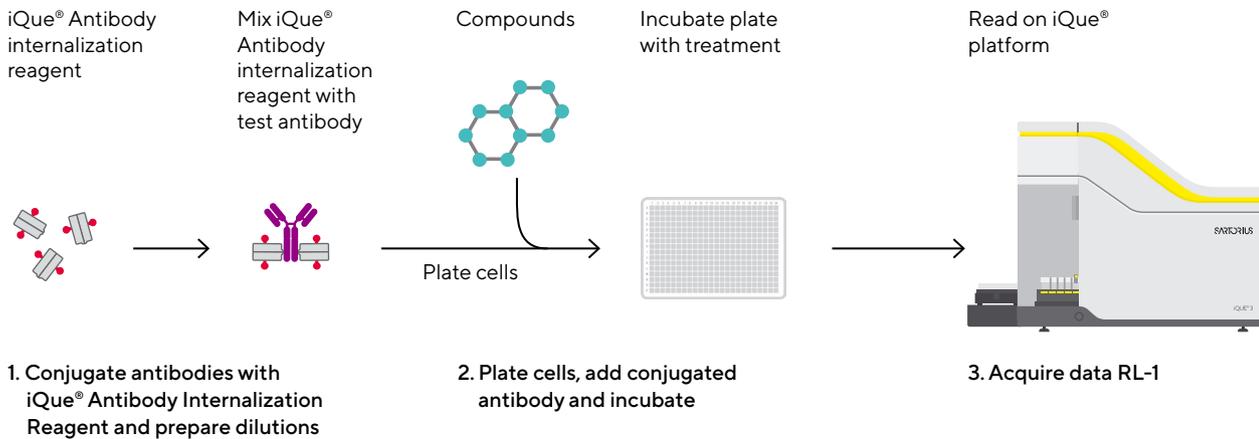


Because the reagent binds to the Fc region of the antibody of interest, it is essential to choose the appropriate type (mouse or human) of the iQue® Antibody Internalization Reagent.

This high-throughput, no-wash assay measures antibody internalization from 20 μ L of sample and can be used to profile a large number of antibodies in a 96- or 384-well format.

Antibodies are labeled with the novel, pH-sensitive iQue® Antibody Internalization Reagent. The labeled antibodies have little fluorescence at neutral pH, but become highly fluorogenic at a lower pH when they are internalized and processed through the acidic lysosome/endosome pathway. After a two hour incubation, sample acquisition is performed using the iQue® platform (with sampling times of less than 5 minutes for a 96-well and 20 minutes for a 384-well format). The integrated iQue Forecyt® software enables data analysis and visualization with plate-level analytics.

Assay Workflow Overview



Reagents Provided

Components	1 × 96-well format (10 µg lyophilized powder)	1 × 384-well format (40 µg lyophilized powder)
iQue [®] Human Antibody Internalization Reagent	Cat. No. 90564	Cat. No. 90565
iQue [®] Mouse Antibody Internalization Reagent	Cat. No. 90566	Cat. No. 90567

Note: Cat. No. 90564 and 90566 (1 × 96-well format) includes 10 µg of iQue[®] Antibody Internalization Reagent. This is enough for 1 × 96-well plate of a single test antibody (at up to 4 µg/mL top concentration, 12 replicates) with overage when reconstituted. Cat. No. 90565 and 90567 (1 × 384-well format) includes 40 µg of iQue[®] Antibody Internalization Reagent. This is enough reagent for 1 × 384-well plate or 4 × 96-well plates with overage when reconstituted.

Detection Channels

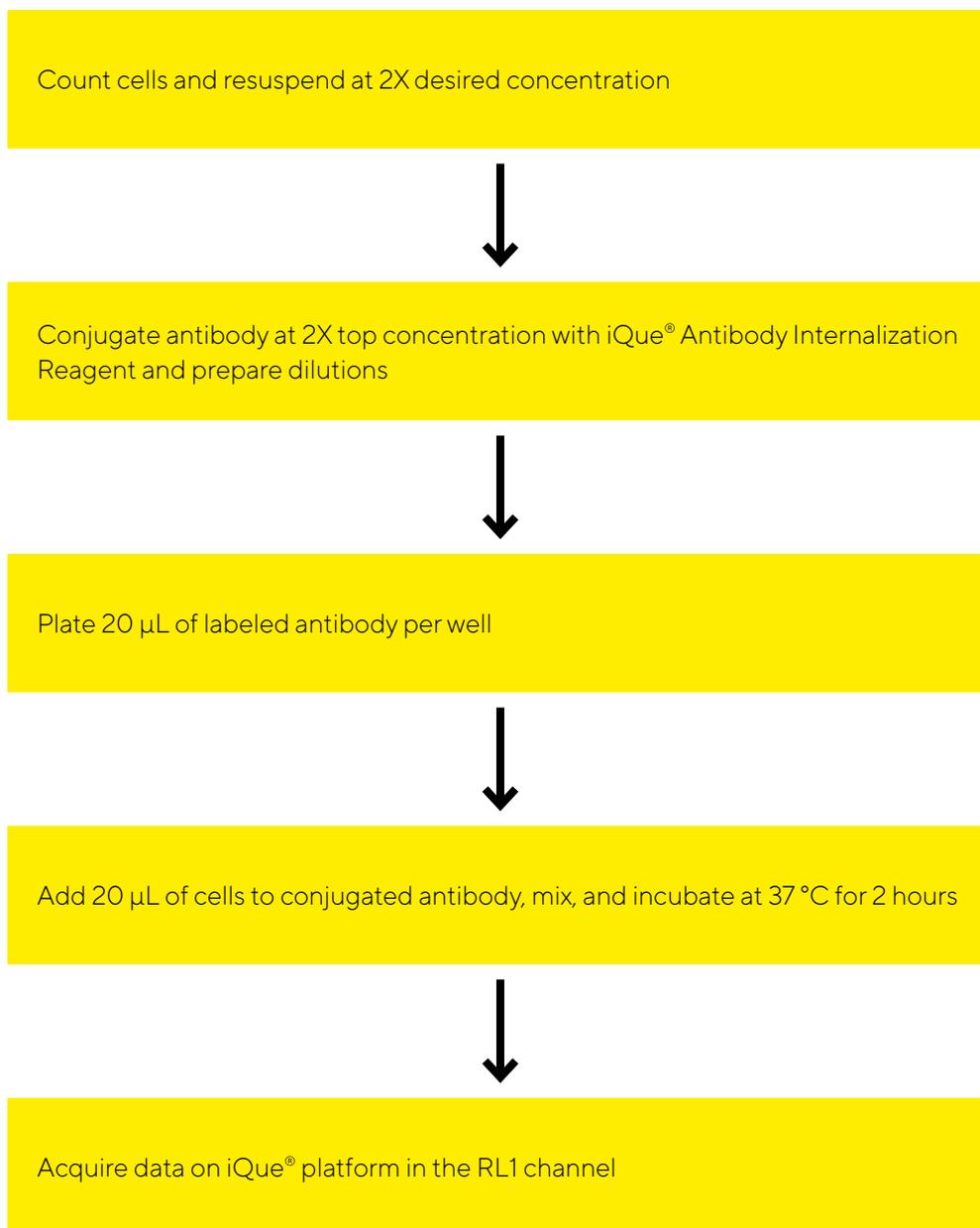
The iQue[®] Antibody Internalization Reagent uses the RL1 detection channel. This reagent was validated for use on the iQue[®] platform.

iQue[®] Screener PLUS Detector Channels (VBR)

Detector	Spectrum	Violet Laser (405 nm)	Blue Laser (488 nm)	Red Laser (640 nm)
445/45 nm		VL1		
530/30 nm		VL2	BL1	
572/28 nm		VL3	BL2	
615/24 nm		VL4	BL3	
675/30 nm		VL5	BL4	RL1 iQue [®] Antibody Internalization Reagent Detection Channel
780/60 nm		VL6	BL5	RL2

Assay Overview

This overview summarizes the protocol for use of iQue® Antibody Internalization Reagents. Detailed instructions are provided in the Protocols and Procedures section. Workflow and volumes are identical for both 1 × 96- and 1 × 384-well formats. For first time assay users, refer to Assay Protocol for detailed step by step procedures. The Assay Overview is a tool you can utilize once you are familiar with the protocol.



Protocols and Procedures

Storage and Stability

iQue® Antibody Internalization Reagent is light sensitive; protect from light.

Store lyophilized product at 2–8 °C. Rehydrate with sterile water and vortex gently until dissolved. Remaining reconstituted reagent can be stored at -80 °C for > 1 year. Avoid repeated freezing and thawing. Antibody labeling should be performed immediately before use.

Recommended Materials

We strongly recommend running a positive and negative isotype control along with the test antibody.

- For the iQue® Mouse Antibody Internalization Reagent, the positive control is human anti-CD71 produced in mouse (Sigma SAB4700520-100UG, supplied at 1 mg/mL in PBS) and the negative isotype control is Mouse IgG1 (R&D Systems, MAB002).
- For the iQue® Human Antibody Internalization Reagent, the negative isotype control is Human IgG1 (Absolute Antibody, AB00178-10.0).

Materials Required but Not Provided

- Test antibody of interest (with human or mouse Fc region) at a known concentration
- Target cells of interest
- Target cell growth media
- Multichannel pipette capable of delivering 20 µL (e.g. Sartorius Tacta or Picus)
- 96-well v-bottom plate (iQue® #10149) or 384-well v-bottom plate (Greiner #781280)

Cell Preparation

Count target cells and adjust concentration with growth media to 2X desired final assay concentration in a volume sufficient to ensure 20 µL per well with overage. We suggest a final assay concentration of 1×10^6 cells/mL.

For a 96-well format, we recommend at least 3 mL of cells prepared at 2×10^6 cells/mL (6 million cells) to accommodate liquid handling.

For a 384-well format, we recommend at least 12 mL of cells prepared at 2×10^6 cells/mL (24 million cells) to accommodate liquid handling.

Preparation of Reagent for 1 × 96- and 1 × 384-Well Format

1. Rehydrate iQue® Antibody Internalization Reagent with 100 µL sterile water for 1 × 96-well format (or 400 µL sterile water for 1 × 384-well format). This results in a final concentration 100 µg/mL for either format. The reagent is light sensitive; protect from light.

Prepare a 1:3 molar ratio of test antibody to iQue® Antibody Internalization Reagent. The reagent is approximately one third the size of an antibody, equal mg/mL quantities will produce a 1:3 molar ratio of test antibody to reagent.

2. Calculate the amount of labeled antibody needed using the following formula:

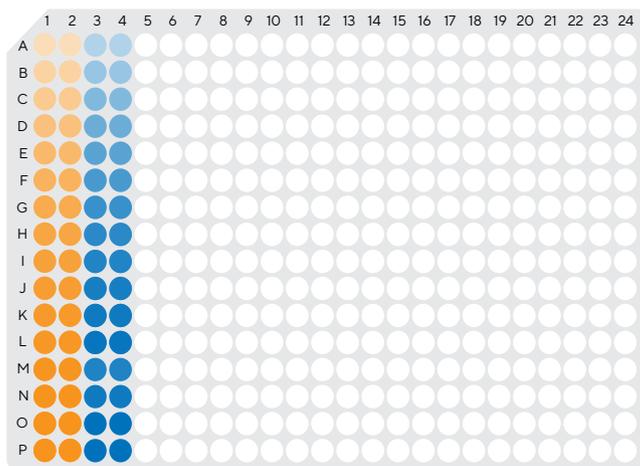
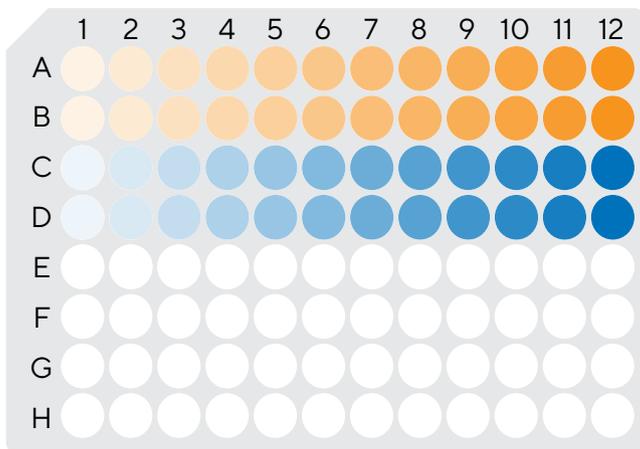
Calculation for 2 Replicates: Mouse Antibody Labeling Using Positive Control Anti-CD71 at 1 mg/mL Stock Concentration for 1 × 96- Well Format

1. Determine final assay concentration of test antibody. 1 µg/mL for anti-CD71 is recommended for a positive control. Prepare working stock at 2X or 2 µg/mL.
2. Determine volume of test antibody required at 2X for 2 replicates of 20 µL per well top concentration as well as adequate overage for 1:2 serial dilutions:
 $(\# \text{ of wells}) \times (\text{volume per well}) \times (\text{overage for serial dilutions})$
2 wells × 20 µL per well 5 = 200 µL labeled antibody at 2X
3. Calculate volumes of test antibody, iQue® Antibody Internalization Reagent, and media for a 2X final assay concentration of labeled antibody:
 - a. Determine volume of test antibody. For a small number of replicates, dilute test antibody stock to 100 µg/mL for more accurate pipetting:
 $(\text{total volume of labeled antibody}) \mu\text{L} \times (2\text{X final concentration of test antibody}) \mu\text{g}/\text{mL} / (\text{diluted stock concentration of test antibody}) \mu\text{g}/\text{mL}$
 $(200 \mu\text{L}) \times (2 \mu\text{g}/\text{mL}) / (100 \mu\text{g}/\text{mL}) = 4 \mu\text{L}$ of test antibody (at 100 µg/mL)
 - b. Determine volume of iQue® Antibody Internalization Reagent:
 $(\text{volume of test antibody}) \mu\text{L} \times (\text{diluted stock concentration of test antibody}) \mu\text{g}/\text{mL} / (\text{stock concentration of iQue® Antibody Internalization Reagent}) \mu\text{g}/\text{mL}$
 $(4 \mu\text{L}) \times (100 \mu\text{g}/\text{mL}) / (100 \mu\text{g}/\text{mL}) = 4 \mu\text{L}$ iQue® Antibody Internalization Reagent
 - c. Determine volume of media:
 $(\text{total labeled antibody volume}) - (\text{test antibody volume}) - (\text{iQue® Antibody Internalization Reagent volume})$
 $(200 \mu\text{L}) - (4 \mu\text{L}) - (4 \mu\text{L}) = 192 \mu\text{L}$ media

Assay Protocol for for 1 × 96- and 1 × 384-Well Format

Label test antibody and prepare dilutions. We strongly recommend running an appropriate positive and negative isotype control along with the test antibody for the mouse or human type.

1. Add test antibody and iQue® Antibody Internalization Reagent (using previous calculations) to culture media in a tube. Protect from light, and incubate for 15 minutes at 37 °C.
2. Prepare 1:2 serial dilutions of the labeled antibody in culture media. This can be done in a separate 96-well format or in tubes.
3. Plate 20 µL of dilutions (including a media-only blank) in a 96- or 384-well plate from low to high concentration. The template includes positive and negative controls run in duplicate.



4. Add target cells to plated, labeled antibody dilutions. Depending on the number of samples and replicates, you may choose to plate either horizontally or vertically.
 - a. Use a multichannel pipette to add 20 µL of well-mixed target cells at 2X to the wells of a 96- or 384-well format.
 - b. Mix using either the plate shake function on the iQue® platform (2000 RPM) or manually with a multichannel pipette. For plate Shaker instructions see Appendix A.
5. Cover plate and incubate at 37 °C for 2 hours.
6. Acquire data on the iQue® platform.

Note: The work flow and volumes are identical for 1x 96- and 1x 384-well formats.

System Operation

1. Launch iQue Forecyt® software.
2. If you have not already imported the template provided by iQue®, import it now, and select the correct format (96- or 384-well). Create a New Experiment using the template.

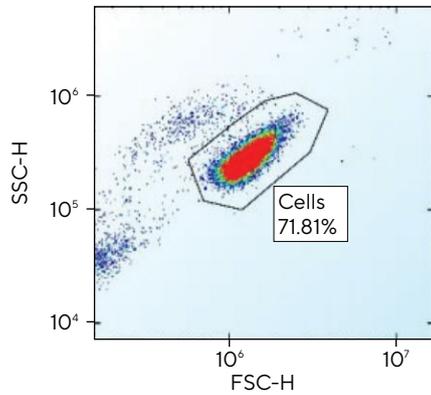
File→New Experiment→Use Template→Experiment Name
3. If you have samples other than the recommended controls on your plate, you can set up Series dilutions in the Series tab. You can also change the concentrations or layout of Series at this time.

Design→Series→Add Series
4. Be sure your plate is in place and click “Run” on the Menu to acquire data.

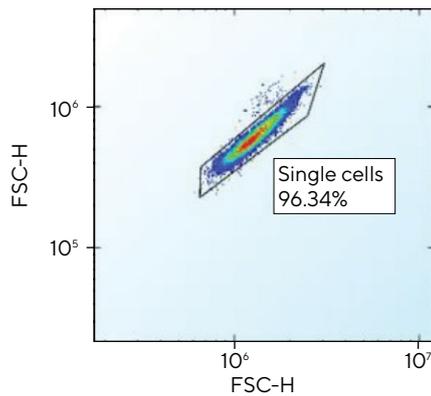
Data Analysis

1. Identify cell populations: the template gates are pre-set for different populations (cells, single cells, and internalized cells). Below are the gating details if you want to manually draw the gates or fine tune the existing gates from the template:
 - a. In all events, FSC-H vs. SSC-H, draw gate around “cells” avoiding cell debris.
 - b. Make a new plot, FSC-H vs. FSC-A, gating on “cells,” and draw another gate around “single cells.”
 - c. Make a histogram based on “single cells,” change the x-axis to “RL1-H,” and draw a gate for “internalized cells” which will appear furthest to the right.

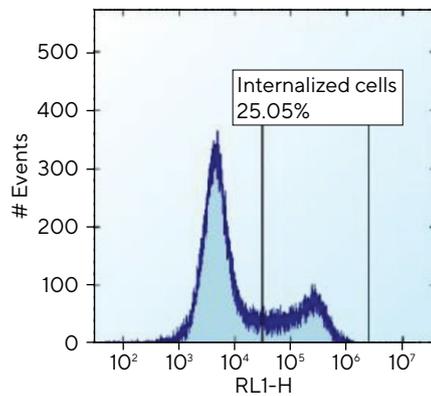
All Events



Cells

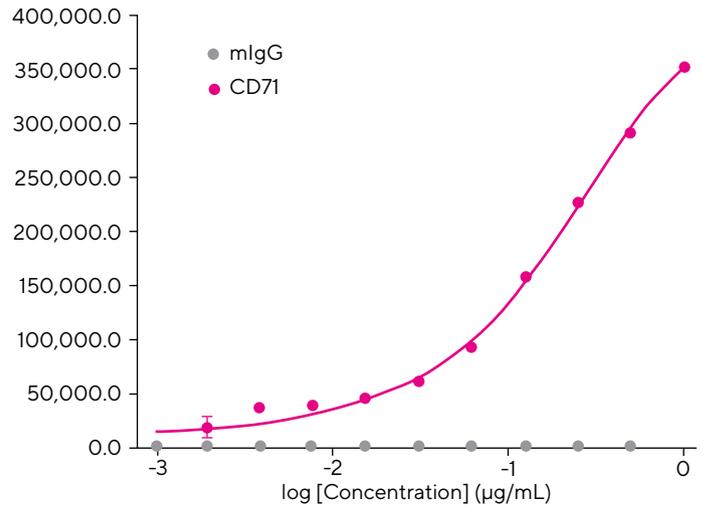


Single Cells

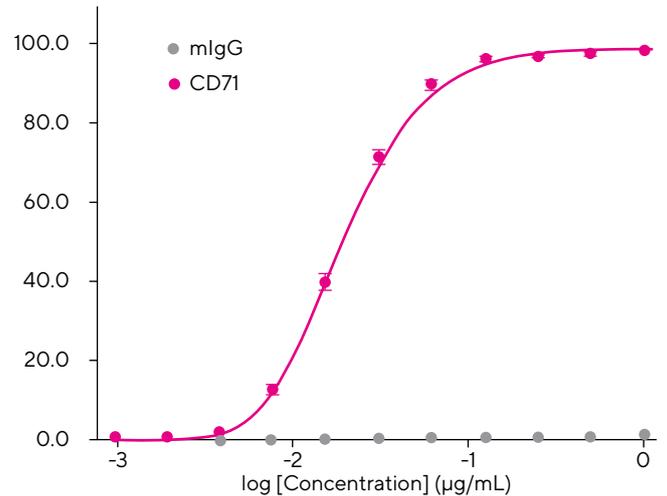


Examples of Positive and Negative Control Antibody Dose Response Curves Showing MFI and Percent Internalized for Jurkat Cells:

Median RL1-H of Internalized Cells



Internalized Cells as % of Single Cells



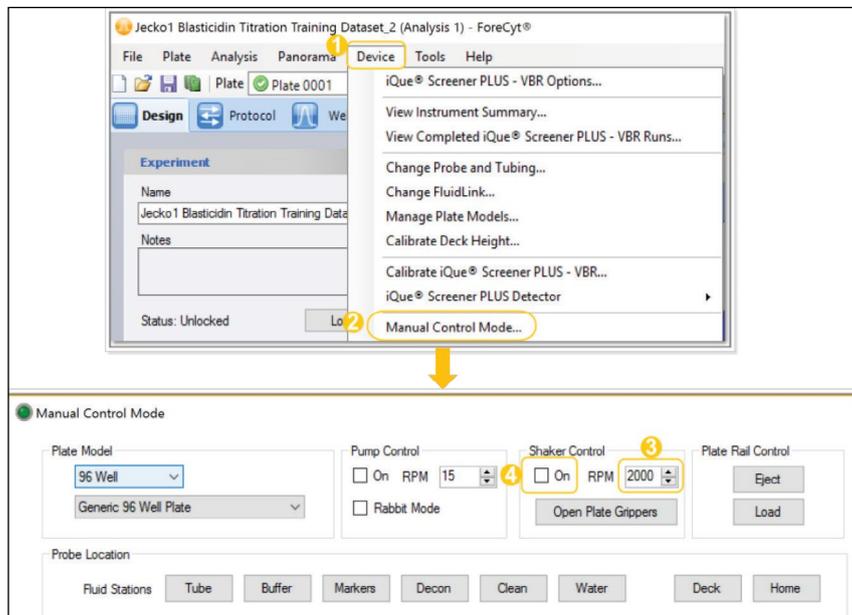
2. Plot dose response curves: the template includes a dose response curve plotted for Mean Fluorescent Intensity (MFI) and percent internalized for a control sample. Follow these steps to create curves for additional samples:
 - a. Create Primary Metric
Metrics→**Add**→**Primary Metric**
(Primary Metrics for "median RL1-H of internalized cells" and for each antibody and/or cell type tested)
 - b. Create Advanced Metric
Metrics→**Add**→**Advanced Metric**
(“Internalized cells as percent of single cells” and for each antibody and/or cell type tested)

Appendix A

Mixing Samples Using Shaker

This assay requires shaking. If you don't have a separate shaker, you can use the one on the iQue® platform.

1. Click on Device in the menu bar.
2. Scroll down to Manual Control.
3. In the Manual Control window, use the arrows to set the RPM.
4. As soon as you click On, the shaker will begin to shake and continue to shake until you unclick.



**For Research Use Only.
Not For Therapeutic or Diagnostic Use.**

North America

Sartorius Corporation
300 West Morgan Road
Ann Arbor, Michigan 48108
USA
Phone +1 734 769 1600

Europe

Sartorius UK Ltd.
Longmead Business Centre
Blenheim Road
Epsom
Surrey, KT19 9QQ
United Kingdom
Phone +44 1763 227400

Asia Pacific

Sartorius Japan K.K.
4th Floor, Daiwa Shinagawa North Bldg.
1-8-11, Kita-Shinagawa 1-chome
Shinagawa-Ku
Tokyo 140-0001
Japan
Phone +81 3 6478 5202