A New Technology for In Vitro Chemotaxis Assays

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Introduction

- · Chemotaxis is the movement of a cell in a direction corresponding to a gradient of increasing or decreasing concentration.
- · Chemotaxis is a fundamental element of normal and pathological cell biology.
- Traditional in vitro methods for studying cell migration include:
 - · Scratch or Cell Exclusion Assays: These are not measures of directed cell migration or chemotaxis. For the most part, they are a measure of "random" migration
 - · Microfluidic Chemotaxis Assays: Researchers can see the cells, but they suffer from small gradients across the cell, low participation rates, and low throughput
 - Traditional Boyden Chamber Assays: This predominant industrial approach has good throughput (96-wells). However, the researcher can not easily visualize the process of cell migration, it requires many cells, and additional labeling or manual cell counting

This poster describes a novel approach that combines hardware, software algorithms, and a consumable to provide a fully automated, integrated solution for studying chemotaxis using live-cell imaging.

IncuCyte ClearView Cell Migration Plate

OOO naced 8 µm pores

Standard Boyden Chambe

stander.

ClearView Cell Migration Plate

24 36 48 60 Time (hr)

Evaluation of the gradient. A 10kD dextran (labeled with Alexa Fluor[®] 594) was added to the ClearView reservoir plate at a concentration of 10 μM to establish gradients

of diffusion were made by sampling the insert wells and measuring fluorescent intensity on a microplate reader. Each data point represents mean ± SEM, N=3.

over zero. 24, 48, and 72 hours. Measu

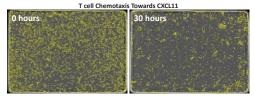


- Visualize Chemotaxis The ClearView Plate incorporates an optically smooth membrane surface enabling acquisition of high-definition, phase-contrast images. Standard Boyder Chamber surfaces are not easily amenable to imaging.
- Persistent Gradient The low porosity of the ClearView Plate results in a gradient that is stable for over 72 hours compared to 4 hours in traditional consumables
- Low Cell Density The combination of a long-term, persistent gradient and the interest in visualizing chemotaxis has resulted in an assay that requires significantly fewer cells compared to traditional Boyden Chamber Assays.
- · Integrin Signaling In the ClearView Plate, cells are required to migrate to the pores. This requires integrin interactions with the substrate that likely are not required in traditional Boyden Chamber consumables
- Automated Image Processing The unique design of the ClearView Plate facilitates quantitation of cells on top and the bottom of the membrane

IncuCyte ZOOM®

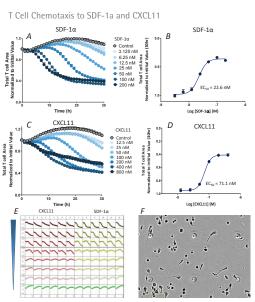


Automated, Label Free Quantitation



The phase-contrast image is blended with image segmentation mask (yellow) created by an automated image processing algorithm.

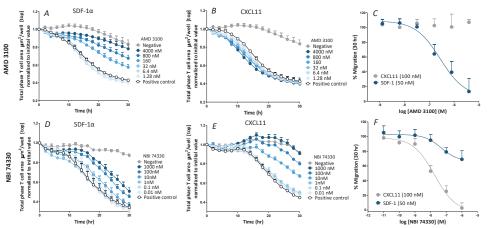
- Optically-clear surface for label-free imaging within IncuCyte ZOOM[®]
- · Cells are added to the upper chamber and chemoattractant to lower reservoir plate.
- · Chemotactic cell migration towards the pores is automatically analyzed using the IncuCyte ZOOM® instrument.



5.000 CD3/CD28 Dynabead-activated T cells were seeded in each well on an ICAM-1 coated surface. The indicated chemoattractant was added to the reservoir plate. Data were collected over a 36 hour period at 1hr intervals. A and C Kinetic curve of concentration-dependent responses to SDF-I α and CXCL11. Data represent mean ± SEM; N=6. B and D: SDF-1α and CXCL11 agonist curve at 30 h Data represents the mean ± SEM; N=6 per condition. E: Each well is individe graphed in a microplate graph overview, illustrating well-ov-well reproducibility. F. A representative image acquired at first time point of the assay from control wells (no chemoattractant). Lamellipodium/Filopodium on T cells indicates that the cell are actively interacting with the surface.

Specific Inhibition of T Cell Chemotaxis using CXCR3 and CXCR4 Inhibitors

5.000 neutrophils per well were seeded on a Matrigel + FBS coated surface. The indicated chemoattractant was added to the reservoir plate. Data were collected over a 6 hour period at 30 min intervals. A and C: Kinetic curve of concentration-dependent responses to IL-8 and fMLP. Data represent mean ± SEM; N=4. B and D: IL-8 and fMLP agonist curve at 4 h. Data represent mean ± SEM; N=4 per condition. E: Each well is individually graphed in a microplate graph overview, illustrating well-to-well reproducibility. F: Representative image acquired at first time point of the assay from control wells (no chemoattractant). Blurry cells are localized on the bottom side of the membrane.



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the cells at indicated concentrations. SDF-1a (Ca0 AMD 3100 and NBI 74330. CD3/CD28 Dynabeads-activated T cells were plated at a density of SK/well on a coated ClearView insert (Protent of +1/cA0). AMD 3100 (**A** and **B**) or NBI 74330. CD3/CD28 Dynabeads-activated T cells were plated at a density of SK/well on a coated ClearView insert (Protent of +1/cA0). AMD 3100 (**A** and **B**) or NBI 74330. CD3/CD28 Dynabeads-activated T cells were plated at a density of SK/well on a coated ClearView insert (Protent of +1/cA0). AMD 3100 (**A** and **B**) or NBI 74330. CD3/CD28 Dynabeads-activated T cells were plated at a density of SK/well on a coated ClearView insert (Protent of +1/cA0). AMD 3100 inbitot has a clear selective effect on CCKH-mediated demonstais towards SDF-1a. (CCG AMD 3100 = 197 nM). No effect of AMD 3100 was found in DCKR3-meadiated chemotaxis towards CXCL11. NBI 74330 has a clear effect on CXCL11-mediated chemotaxis (CSO NBI 74330 = 17.8 nM). NBI 74330 weaky inhibited chemotaxis towards SDF-1a.

Summary and Impact

- Real-time visualization and automated analysis of chemotactic cell migration in a 96-well format within your incubator
- Measure label-free, or labelled cell migration with fixing, staining or cell scraping steps
- Setup and walk away fully automated image based analysis
- Highly reproducible 96-well approach suitable for profiling and screening
- Investigate cell migration on biologically relevant surfaces
- Sustained and stable gradient over 72 hours





4.6 nN 13.8 nN • 41.5 nN

124 nM

1120 nM

fMLP

Contro

1.37 nM

4.12 nM

12.4 nM

37.0 nM

111 nM 🗣 333 nM

1000 nM



EC ... = 24.2 nN

EC₁₀ = 7.56 nM

-8 -/ Log [IL-8] (M)

Log (fMLP) (M)

D