SARDRIGS

Simplifying Progress

Optimization of the HEK293T Suspension Cultivation With a DoE-Approach in Ambr[®] 15 Cell Culture

Franziska Bollmann¹, Diana Riethmüller¹, Erik Johansson², Alexander Tappe¹

¹R&D Regenerative Medicine, Sartorius Stedim Biotech, Göttingen, Germany ²Sartorius Stedim Data Analytics AB, Sweden. July 2019 Royston-Info@sartorius.com

Introduction

• HEK293T cells are commonly used as a workhorse cell line for viral vector production for cell and gene therapy applications.

Materials and Methods

 Adherent HEK293T cells have been adapted to suspension culture in serum-free conditions



- A significant challenge for the Regenerative Medicine (RM) industry is to develop
- a HEK293T suspension cell culture process that is well characterized and can be scaled up for production to ensure clinical and commercial success.
- Ambr[®] 15 Cell Culture is an automated micro-scale bioreactor system that mimics the features and process control (pH, DO, temperature, stirring rate) provided by much larger scale bioreactors, but in a volume of 10 - 15 mL. Parallel processing capability and excellent consistency enable rapid, high throughput process improvement and optimization, including DoE studies.
- High throughput tools with parallel processing, such as Ambr[®] 15, help to address a major manufacturing bottleneck. They can be used as a scale-down model for process development, clone selection and effective media optimization in less time with reduced reagent use and labor saving.
- Design of Experiments (DoE) is a rational and cost-effective approach to practical experimentation that allows the effect of variables to be assessed using only the minimum of resources. MODDE[®] is a state-of-the-art DoE software package. It enables fast and effective identification of critical process parameters (CPPs) and, subsequently, establishment of a Design Space, resulting in reduced bioprocess complexity and increased process understanding.

Scope of Work

• Use Ambr[®] 15 for HEK293T suspension culture optimization aiming for VCC (viable cell count/density), viability and generation time to be comparable to standard shake flask culture: identify optimum stirring speed, DO value and pH value. Perform a DoE study to identify optimal culture conditions by using MODDE[®] software for experimental planning and analysis of results.

Results: Experiment 1

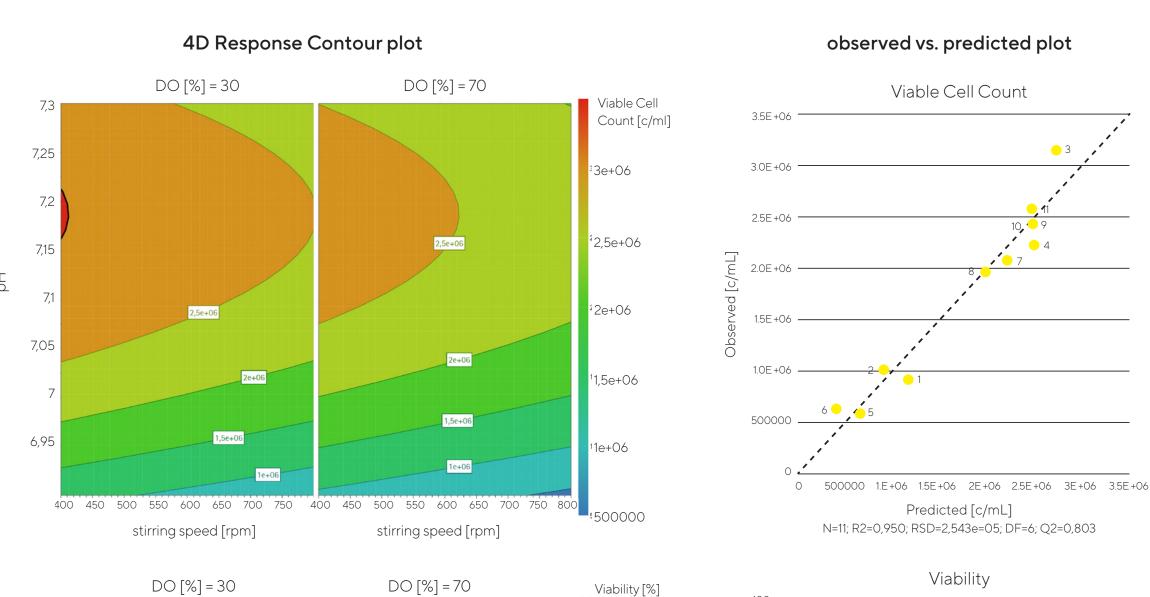
DoE study for optimization of HEK293T cultivation

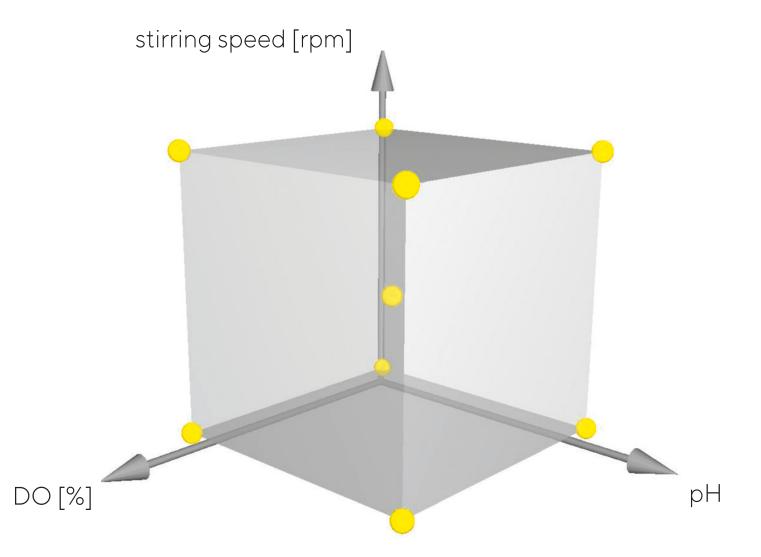


- optimal culture medium was CD293 + 4mM Glutamax (Gibco) as determined in a previous study
- Setpoints Ambr[®] 15 culture using sparged vessels:
- bioreactor temperature: 36.8°C
- inoculation density: 3x105 cells/mL
- fill volume: 15 mL, inoculum volume: 2 mL
- daily antifoam c addition
- with MODDE[®] software a 2-level full factorial design with three centerpoints was used for setting up a DoE (experiment 1)
- setpoints shake flask:
- incubator temperature: 36.8°C
- inoculation density: 3x105 cells/mL
- shaking rate: 120rpm (baffled flask)
- fill volume: 37.5mL
- CO²: 8%
- orbit: 5cm

Table 1: overview of process parameters and readouts of the DoE study

Range				
400	800			
6.9	7.3			
30	70			
VCC, viability, generation time				
	400 6.9 30			

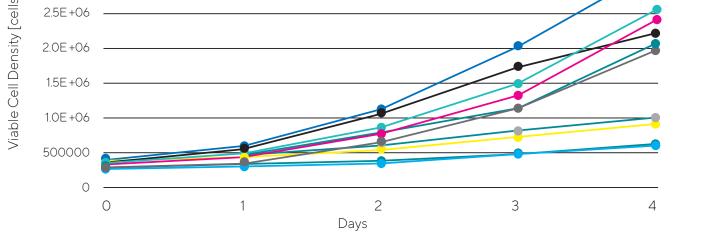




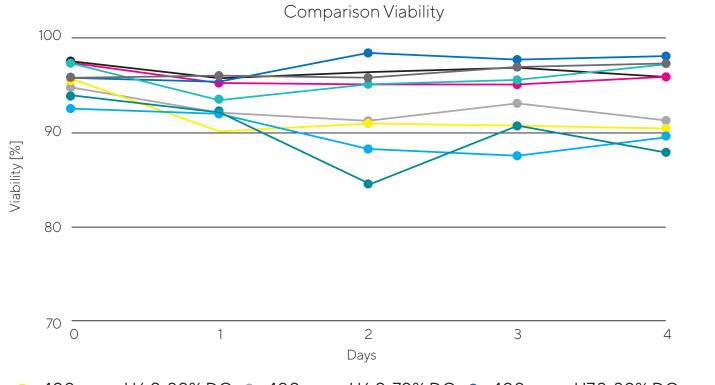
Results: DoE follow up Experiment 2

Comparison of Ambr[®] 15 with shake flask culture

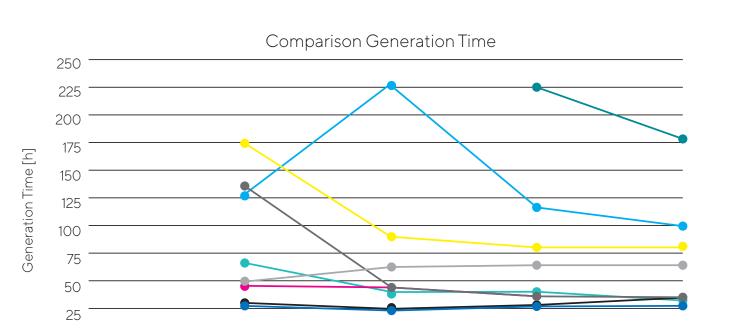
- selected conditions: analyzed in duplicates • 2 stirring speeds: 300 and 400 rpm and 4 pH values: 7.1, 7.2, 7.3, 7.4 • cultivation in the Ambr[®] 15 yields increased viable cell count compared to standard shake flask



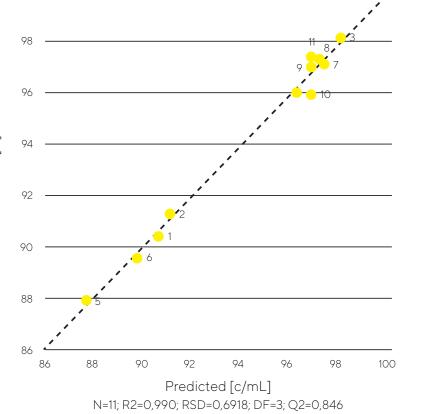
← 400rpm; pH6.9; 30% DO — 400rpm; pH6.9; 70% DO — 400rpm; pH7.3; 30% DO ← 400rpm; pH7.3; 70% DO ← 600rpm; pH7.1; 50% DO ← 600rpm; pH7.1; 50% DO ← 600rpm; pH7.1; 50% DO ← 800rpm; pH6.9 30% DO ← 800rpm; pH6.9 70% DO



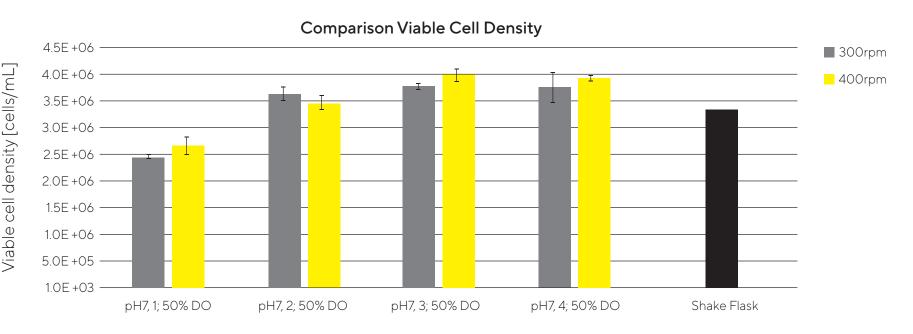
← 400rpm; pH6.9; 30% DO — 400rpm; pH6.9; 70% DO — 400rpm; pH7.3; 30% DO ← 400rpm; pH7.3; 70% DO ← 600rpm; pH7.1; 50% DO ← 600rpm; pH7.1; 50% DO ← 600rpm; pH7.1; 50% DO ← 800rpm; pH6.9 30% DO ← 800rpm; pH6.9 70% DO

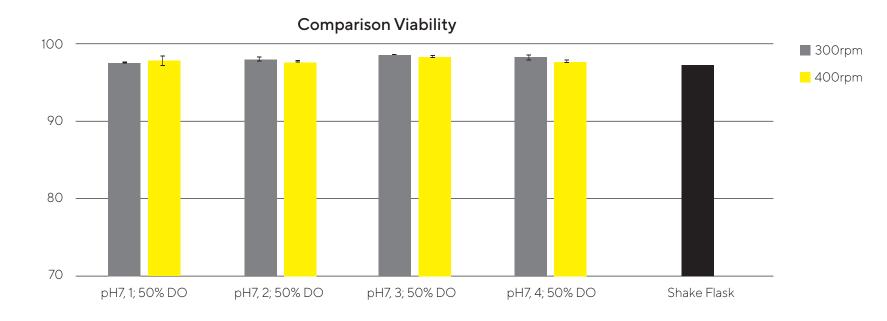


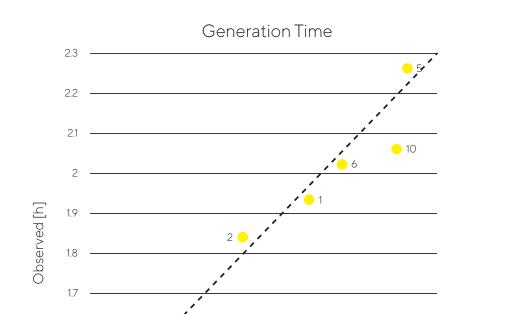
N=11; R2=0,950; RSD=2,543e=05; DF=6; Q2=0,803

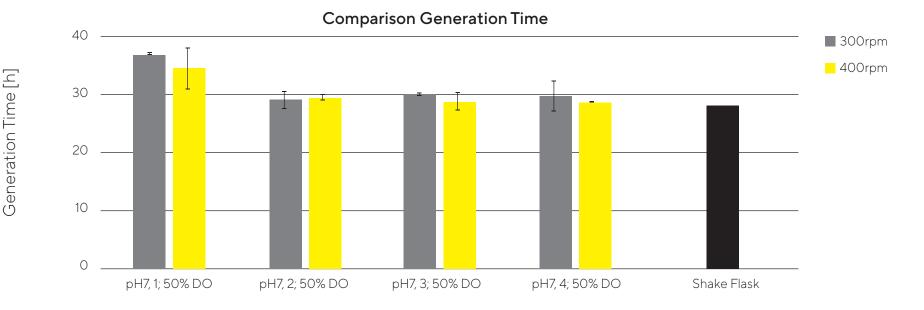


Ambr[®] 15 performs equal to shake flask wrt generation time viability of cells is generally better in Ambr[®] 15 than in shake flask • overall high level of reproducibility between replicate vessels observed PH 7.3 at 400 rpm stirring speed determined to be optimal for cell growth generation time and VCC are dependent on pH value and stirring speed



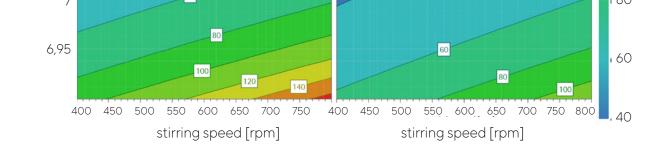








← 400rpm; pH6.9; 30% DO — 400rpm; pH6.9; 70% DO — 400rpm; pH7.3; 30% DO ← 400rpm; pH7.3; 70% DO ← 600rpm; pH7.1; 50% DO ← 600rpm; pH7.1; 50% DO ← 600rpm; pH7.1; 50% DO ← 800rpm; pH6.9 30% DO ← 800rpm; pH6.9 70% DO



400 450 500 550 600 650 700 750 800

stirring speed [rpm]

Generation

Time [h]

160

140

120

100

DO [%] = 70

1.5 —	<u> </u>	78						
1.4	,							
1.4	1.5	1.6	1.7	1.8m	1.9	2	2.1	2.2
				Predict	tod [h	1		

Response	Ambr [®] 15	Shake flask
VCC [cell/mL]	4.01x10 ⁶	3.35x10 ⁶
Viability [%]	98.4	97.5
Generation time [h]	28.8	28.1

Results: Find optimal settings within design space to fulfill acceptance criteria:

7,25

7,05

6,95

7,25

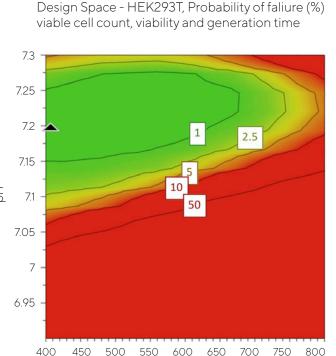
7,05

400 450 500 550 600 650 700 750

DO [%] = 30

stirring speed [rpm]

- DoE evaluation in MODDE[®] showed a good model validity
- pH and stirring speed were identified to be significant parameters
- optimal setpoint: stirring speed 400 rpm; pH 7.22; DO=30% according to Optimizer function in MODDE[®]
- according to contour plot: possibility that stirring speed <400 rpm could yield better cultivation results
- since DO was not found to be a significant parameter we choose 50% DO as a setpoint for further optimization studies



stirring speed [rpm] Interval = Confidence Acceptance limit = 5% DO:

Key process attributes	Unit	Optimization Objective	Min	Target	Max		Prob. of aliure	Predicted output at optimum
Viable cell count	c/ml	Maximize	2.00E+06	3.00E+0)6 -	C).02%	2.9E+06
Viability	%	Maximize	95	100	-	C).17%	98.0
Generation time	h	Maximize	-	20	40	C).29%	29.9
Process paramet	er Unit	Role	Value L	ow limit	High limit		Optimal	settings
DO	%	Constant	50 -		-	!	50	
рН		Free	- 6	5.9	7.3		7.2	
Stirring speed	rpm	Free	- Z	100	800		400	

Summary and Outlook

This study demonstrates that the Ambr[®] 15 Cell Culture system in combination with the DoE MODDE[®] software enables a systematic investigation of critical process parameters and rapid, high throughput process improvement and optimization. The results prove that the transition from shake flask to a scalable stirred bioreactor system can be accomplished very fast. A key next step is to use the identified HEK293T culture conditions to perform a DoE study with the Ambr[®] 15 to optimize viral vector production for cell and gene therapy applications.

Process parameters	Optimized cultivation setpoints Ambr® 15
Stirring speed [rpm]	400
рН	7.3
DO [%]	50