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Protocol

Detailed Incucyte® Chemotaxis Assay Protocol for Jurkat Cells

The following protocol is a detailed example designed to enable you to run a successful Incucyte[®] Chemotaxis Jurkat Migration Assay. We provide three membrane coating options which allow for either clustered or single cell migration.

Materials

- Jurkat (ATCC, TIB-152)
- Unopened bottle of RPMI 1640 (Life Technologies 11875-085)
- Fetal Bovine Serum (FBS; Sigma-Aldrich F2442)
- Matrigel[®] (Corning 354234)—optional
- Fibronectin (Sigma Aldrich, F1141)
- Protein G (Life Technologies 101200)-optional
- ICAM (Life Technologies 10346-H03H or Sino Biologicals Inc. 10346-H03H)—optional
- Bovine Serum Albumin (BSA; Sigma Aldrich, A7906)
- D-PBS (w/o Ca²⁺, Mg²⁺, Life Technologies 10010)
- SDF-1a (R&D Systems 350-NS-050)
- Incucyte[®] Clearview 96-well Plate for Chemotaxis (Sartorius Cat. No. 4582, 4599, or 4648)

Membrane Coating Options

Coating with Matrigel®

Clustered Cell Migration

 Pre-cool an Incucyte[®] Clearview Plate in a CoolBox[®] system containing a frozen cold pack and CoolSink[®] plate (4 °C), for 5 minutes.

Note: It is important to keep close temperature control of Matrigel[®] to prevent unwanted gelling. The Incucyte[®] Cell Invasion Kit (Sartorius Cat. No. 4444) includes a specialized CoolBox[®] system to ensure the temperature of your assay plate and biomatrix materials are maintained between 4-8°C-preventing premature polymerization and eliminating edge effects. Crushed ice can be used as an alternative, however non-uniform cooling can lead to assay variability.

- 2. On ice, prepare 50 μg/mL Matrigel[®] + 10% FBS diluted in pre-chilled RPMI + 0.5% FBS. Gently invert to mix.
- 3. Using reverse pipetting, add 150 μ L of the Matrigel[®] solution to the chilled reservoir plate wells. At a slight angle, place the insert plate into the reservoir plate and gently roll the plate into position. Using reverse pipetting, add 20 μ L of the Matrigel[®] solution to the insert wells.

- 4. Place the plate at 37 °C and incubate for 30 minutes.
- Remove the Incucyte[®] Clearview Plate from 37 °C and allow to cool down to ambient temperature for 30 minutes.

 $\ensuremath{\textbf{Note:}}$ This step is important in order to achieve even cell distribution.

 Prior to cell seeding, aspirate the Matrigel[®] coating from the insert well and reservoir wells of the Incucyte[®] Clearview Plate. To the reservoir wells, aliquot 200 μL of D-PBS and gently return the insert into to the reservoir plate.

Note: Alternatively, if removal of Matrigel[®] is not desired, cells can be seeded directly into the wells containing coating. The volume of cells being added to the insert must be reduced to 40 μ L (refer to step 4, Chemotaxis Assay).

Coating with ICAM

Single Cell Migration

- 1. Resuspend Protein G and ICAM reagents in sterile water.
- Using reverse pipetting, coat the top surface of the Incucyte[®] Clearview Plate membrane with 20 μL of 20 μg/mL Protein G solution and incubate for 1 hour at 37 °C.
- Wash the membrane once with D-PBS by adding 40 µL D-PBS directly to the wells containing Protein G. Remove the full volume (~ 60 µL) and promptly proceed with the ICAM coating step.
- 4. Using reverse pipetting, add 20 μL of 5 μg/mL ICAM to the insert wells and incubate for 2 hours at 37 °C.
- 5. Aspirate ICAM from the insert, then block both sides of the membrane with D-PBS + 1% BSA by adding 20 µL to the insert wells and 150 µL to the reservoir wells of the Incucyte®Clearview Plate (pre-fill reservoir and gently place the insert into the reservoir plate containing BSA). Incubate at ambient temperature for 30 minutes.
- 6. After incubation, transfer the insert plate to a new Incucyte[®] Clearview Reservoir Plate containing 200 μL D-PBS in each well. Immediately prior to cell addition, wash the insert wells once with D-PBS by adding 40 μL D-PBS to the insert wells containing 1% BSA, then remove the full volume (~ 60 μL).

Coating with Fibronectin

Single Cell Migration

- 1. Prepare fibronectin at 5 μg/mL in D-PBS (without calcium or magnesium) supplemented with 0.1% BSA.
- Using reverse pipetting, add 150 µL of fibronectin solution into the reservoir of the Incucyte[®] Clearview Plate. Place the insert into the reservoir and, using reverse pipetting, add 20 µL of the fibronectin solution into the insert. In this case, a second reservoir plate will be loaded with chemoattractant and used during the experiment.

- 3. Incubate for 1 hour at ambient temperature.
- 4. Aspirate the fibronectin + 0.1% BSA coating from the reservoir wells and replace with 200 μ L of D-PBS and gently return the insert into to the reservoir plate.
- To the insert, add 60 μL of D-PBS to the wells containing fibronectin + 0.1% BSA, then aspirate immediately prior to cell seeding.

Protocol

Incucyte[®] Chemotaxis Assay

- 1. Thaw Jurkat cell line and wash 1X in 5 mL of serum free RPMI 1640 media.
- 2. Centrifuge cells at 500 x g for 5 minutes.
- Re-suspend cells in an appropriate volume of chemotaxis assay media (newly opened RPMI + 0.5% FBS) and perform a cell count.
- Using a manual multi-channel pipette and reverse pipetting technique, seed cells (60 μL per well, 5,000 cells per well) into every well of the insert plate of the Incucyte[®] Clearview Plate.

Calculation: 83,333 cells/mL x 0.06 mL = 5,000 cells per insert well.

- 5. Allow the Jurkats to settle at ambient temperature on a level surface for 45–60 minutes.
- 6. During cell settling, prepare chemoattractant dilutions and controls (for reference, standard range of SDF-1α is 12.5–125 nM).
- 7. Using a manual multi-channel pipette, add 200 μL of the chemoattractant and control medium to the appropriate wells of the second reservoir plate.
- 8. Carefully transfer the insert plate containing the cells into the pre-filled second reservoir plate containing medium ± chemoattractant.
- 9. Place the Incucyte[®] Clearview Plate into the Incucyte[®] Live-Cell Analysis System and allow the plate to warm to 37 °C for at least 15 minutes.

Note: After 15 minutes, wipe away any condensation that remains on the outside of the plate lid or bottom of the reservoir.

10. In the Incucyte[®] software, schedule 24 hour repeat scanning (10X) for every 30 minutes.

Note: This schedule is only for a scanning a single plate. Fewer scans will be required if scheduling multiple plates.

a. Objective: Ensure 10X objective is installed

b. Vessel Type: Select "Incucyte® Clearview Plate"

c. Channel Selection: Select "Phase"

d. Scan Mode: Select "Chemotaxis (Top/Bot)" scan type and desired Scan Pattern

e. Note the Incucyte[®] Live-Cell Analysis System estimates a scan time of 20 minutes per plate (phase only); however, the actual scan time can take longer.

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