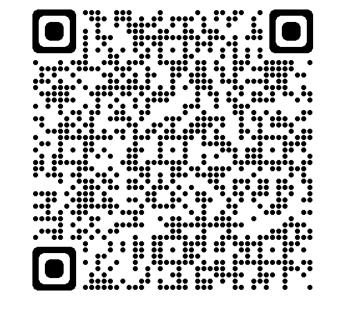
SARDRIG

Simplifying Progress

Unveiling Process Solutions For Plasmid DNA Fermentation Across Upstream Scales

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Introduction

Developing pDNA processes depend upon performant bioreactors to allow a rapid scale up to commercial batches. For this it is relevant to minimize any possible risks while developing a process that fits the industry quality standards. The choice of a well characterized system plays an important role from R&D through to production stages. With the Sartorius Process Solutions portfolio it is possible to accelerate development timelines and ensure process SUCCESS.

Process Characterization with the DECHEMA Guidelines

Figure 1

DECHEMA Characterization Principles

Process Engineering Characterization



Process Engineering Characterization

Define the best parameters to work with (1): Stir speed

That is the reason why Sartorius and JAFRAL partnered together for plasmid DNA production. JAFRAL is a world's leading CMO and CRO for production of bacteriophages and other biomolecules as pDNA. JAFRAL delivers non-GMP and GMP pDNA for various applications, including gene therapy and cell therapy. JAFRAL's GMP facilities enable short turn-around times and best production prices.

The aim of this joint poster with JAFRAL is to provide evidence to demonstrate the benefits of a pDNA process developed using Sartorius scalable solutions. The method chosen to showcase the bioreactor consistency is based on the DECHEMA Guidelines for Engineering Characterization principles. Alongside a strong bioreactor process characterization for pDNA, Sartorius analytical solutions provide the accurate and relevant in-process knowledge for all pDNA production needs.

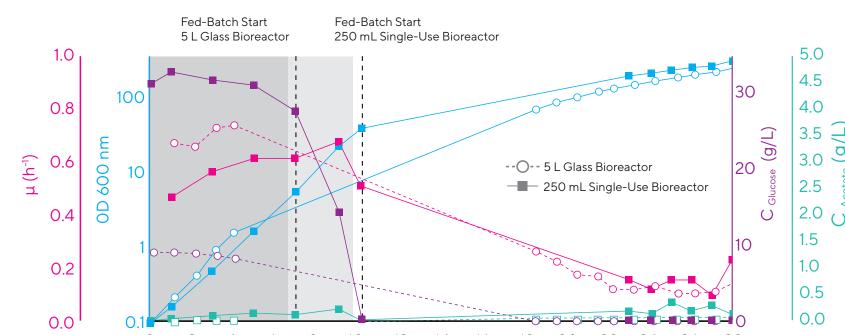
Ambr[®] Platform for Enhanced Optimization

Key biological characterization results on Ambr[®] 250 Modular

- Excellent OTR and mixing support a range of high density cultures
- Comparability proven to 5 L benchtop Universel[®] and larger scale volumes (Table 1)

Figure 2

E. Coli Cultivation in an Ambr[®] 250 Modular System

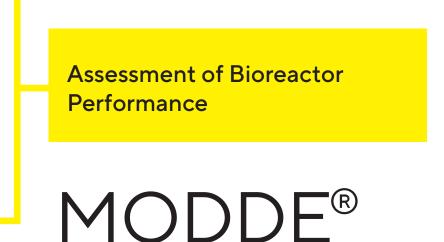


kLa (Gassing out Method), Mixing Time, P/V

DECHEMA 2016 Experimental Methods guidelines defined for Bioreactors (Single-use and Reusable systems) (1)

Microbial/Biological Characterization kLa (Oxygen Balance Method)

DECHEMA Standardized Batch Fermentation Biological Model based on *E. coli* W3110 culture (2) (3)



und Biotechnologie e.V.

These characterizations can benefit significantly from MODDE[®] - Umetrics' suite for enhanced DOE investigations.

Reliable Scale-Up at JAFRAL with Biostat STR[®] Microbial System JAFRAL

Good consistency demonstrated between 15 L stainless steel (SS) and 40 L single-use (SU) and systems

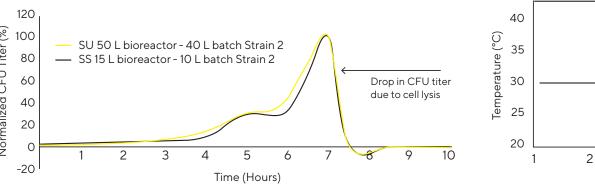
Figure 3B

Rapid scale-up with reduced risk to project timelines

Figure 3A

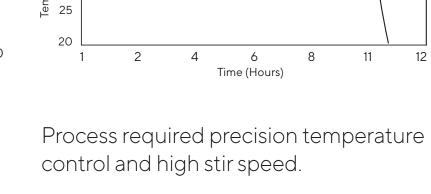






Yield of the process was as expected and comparable with stainless steel fermenter systems.





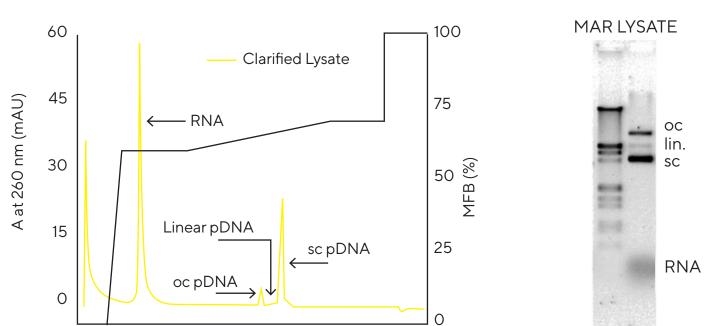
Fast temperature rise in the bag followed

- Maximum working volume
- Mixing times via conductivity/ decolourization
- Power input based on the vessel and motor geometry/torque
- pO_2 with good sensor response time (11s)
- kLa via gassing-out method (1)
- Streamlined Analytical Capabilities
- pDNA characterization with fast and innovative analytics to maximize process productivity:
- CIMac[™] pDNA analytical columns are based on the monolith technology, especially developed for pDNA applications and ideal in the context of process analytical technology (PAT).
- The PATfix[™] HPLC system gives you "at-line" analysis for the control of impurities and critical quality components using HPLC fingerprinting
- These technologies allow you to achieve rapid, high-resolution and flow-independent separations in a matter of minutes.

Figure 4

Figure 5A

Chromatogram Graph (In-House Data) Using CIMac™ pDNA Analytical Column



4 6 8 10 12 14 16 18 20 22 24 26 28

Time (Hours)

Note. Ambr[®] 250 Modular run (squares) compared with a Univessel[®] Glass 5 L reactor (circles); optical density at 600 nm (blue), growth rate (dark pink), acetate (teal), and glucose concentration (purple). Dashed line indicates feed

by fast temperature drop were required to achieve optimal conditions for induction of product synthesis in the fermentation broth.

Temperature control provided was excellent. An increase to 42 °C was achieved within 15 min.

10 15 20 25 Time (Minutes)

PATfix[®] HPLC and CIMac[™] Analytical Columns

Figure 5B

CIMac[™] Analytical Column for pDNA In-Process Control

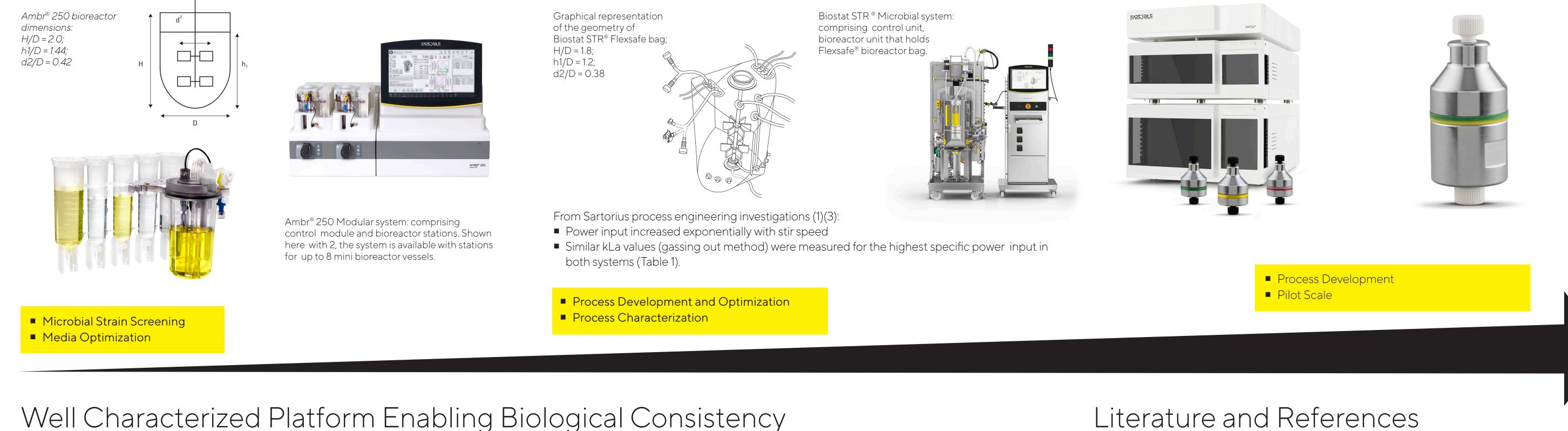


Table 1

Results in Ambr[®] 250 Modular and Biostat STR[®] 50 L

Batch Growth $k_{L}a$ -Value (h^{-1}) Oxygen Fed-Batch Growth Final Fed-Batch References Volume Tip Speed Gas Flow k_La-value (h⁻¹) Mixing (Gassing-Out Method) Times (s) rate³ (μ) (h⁻¹) Balance Method (3) OD 600 nm (L) (m/s)Rate (µ) (h⁻¹) rate (vvm)

1 W. Meusel, C. Loffelholz, U. Husemann, T. Dreher, G. Greller, J. Kauling, D. Eibl, S. Kleebank, Bauer, R. Glockler, P. Huber, W. Kuhlmann, G. T. John, S. Werner, S. C. Kasier, R. Portner, M. Kraume, Recommendations for Process Engineering Characterization of Single-Use Bioreactors and Mixing Systems by Using Experimental Methods, DECHEMA, Frankfurt am Main 2016.

				(
Biostat STR® 50L	40	3.8	1.5	675	<2	0.4 - 0.65	500 - 707*	0.15	~ 300 (<i>E.coli</i> W3110)	Internal data (8)
Ambr [®] 250 Modular	0.25	4.4	1.0	400 ± 7 * 1488 ± 40 **	< 2	0.4 - 0.65	782 ± 27	0.15	~335 (<i>E. coli</i> W3110)	(4) and Internal data (3)

Note. Data sources from literature references, internal studies and data kindly shared from biopharma customers.

* Head Space Exchange term (HSE) not included. **Head Space Exchange term (HSE) included.

Discussion

- Sartorius single-use bioreactors are based on the classic stirring impeller design and have proven mixing times and kLa values being relevant for microbial bioreactors used in industrial processes (3)(4)(5)
- The studies show the reliability and consistency when scaling up with the Ambr[®] and Biostat[®] platforms.
- It has been demonstrated that the Biostat STR[®] Microbial is suitable for *E. coli* cultivation producing plasmid DNA. The process can be scaled-up from Ambr and transferred from stainless steel fermenters.
- Growth data (μ) and maximum produced biomass (OD 600 nm) are reproducible across scales for both batch and fed-batch modes allowing a fast and optimal process development for your cell line as well as flexibility when developing your own platform.
- Higher yields per process can be achieved in a standardized manner.

■ Across all scales biological kLa is minimum of >675 h-1.

- Therefore a biological model initially developed with the Ambr[®] platform can easily be transferred to the larger scale Biostat STR® Microbial including intensified processes with high cell density and requiring higher gassing exchanges.
- The CIMac[™] Analytical technology enables the separation of all three pDNA conformations and can monitor the removal of other impurities such as RNA (6)

Conclusion

- The Ambr[®] platform brings a high throughput strategy for multi-parallel experiments with state-of-the-art automation, fast set-up and high performance especially for R&D and Process Development (4)(5)
- The Biostat[®] STR Microbial based on the GMP ready stand-alone automation platform Biobrain[®] is a reliable option to scale up processes that meet reproducible results at high industry standards.
- Both Ambr[®] and Biostat[®] platforms contribute for well characterized processes and reduced risk during scale up and tech transfer stages alongside strong analytical capabilities with PATfix® HPLC and CIMac[™] Analytical Columns.
- In the recent years plasmid DNA became a product of strongest interest. Sartorius bioreactors demonstrate excellent and easy scalability for this product type from fast process development towards small-scale production helping to enter the market with the highest pace.
- Sartorius brings added value to customers seeking robustness and flexibility in fast paced environments.

- 2 Schirmer, Cedric; Blaschczok, Katharina; Husemann, Ute; Leupold, Marco; Zahnow, Christian; Rupprecht, Jens; Glöckler, Rainer; Greller, Gerhard; Pörtner, Ralf; Eibl-Schindler, Regine; Eibl, Dieter; 2017. Standardized qualification of stirred bioreactors for microbial biopharmaceutical production processes Chemie Ingenieur Technik. DOI: 10.1002/ cite.201700039
- Internal Sartorius studies led by Dr. Marco Leupold (marco.leupold@sartorius.com). 3
- 4 M. Leupold, T. Dreher, M. Ngibuini, G. Greller. A Stirred, Single-Use, Small-Scale Process Development System, Evaluation for Microbial Cultivation. BioProcess International, November 2017.
- 5 Velez-Suberbie, M. L., Betts, J. P. J., Walker, K. L., Robinson, C., Zoro, B. and Keshavarz-Moore, E. (2017), High throughput automated microbial bioreactor system used for clone selection and rapid scale-down process optimization. Biotechnol Progress. DOI:10.1002/btpr.2534
- 6 Application note: In-Process Control of pDNA Production on CIMac[™] pDNA Analytical Column